



OCTO Health RT-PCR Cardiovascular Disease Panel Kit 1

Catalog

RT-CVD-P1-25

RT-CVD-P1-50

Description

Real-Time PCR Cardiovascular Disease Panel Kit 1, 25 reactions

Real-Time PCR Cardiovascular Disease Panel Kit 1, 50 reactions

IVD



Introduction

OCTO Health REAL TIME PCR Cardiovascular Disease Panel Kit 1 is capable of diagnosing a total of fifteen parameters in six tubes. Table below displays the tube numbers parameter locations and which emission channels will be used.

Tube	Gene	Mutation	Rs #	Emission Channel
1	Factor V LEIDEN	G1691A	RS6025	FAM
	Prothrombin	G20210A	RS1799963	HEX
	MTHFR	C677T	RS1801133	ROX
	MTHFR	A1298C	RS1801131	CY5
2	PAI	4G/5G	RS1799762	CY5
	Factor XIII	V34L	RS5985	ROX
3	HPA1	L33P	RS5918	CY5
	β-Fibrinogen	-455G-A	RS1800790	ROX
4	Factor V Cambridge	c.1001G>C	RS118203906	CY5
	H1299R	c.3980A>G	RS1800595	ROX
5	APOB	R3500Q	RS5742904	HEX
	ACE	I/D	RS1799752	CY5
	LTA	804 C>A	RS1041981	ROX
6	Apo E	c.388T>C	RS429358	ROX
	Apo E	c.526C>T	RS7412	CY5

Kit Contents

	CONTENT	25 REACTIONS	50 REACTIONS
1	Master Mix 1	450 µl	900 µl
2	Master Mix 2	450 µl	900 µl
3	Master Mix 3	450 µl	900 µl
4	Master Mix 4	450 µl	900 µl
5	Master Mix 5	450 µl	900 µl
6	Master Mix 6	450 µl	900 µl
7	Control Mix*	180 µl	360 µl

*The Control Mix contains synthetic wild-type and mutant sequences for each parameter listed in the table, except for APOB. APOB includes only the wild-type synthetic sequence. During melt-curve analysis, APOB is expected to yield a wild-type profile, whereas all other parameters should display heterozygous profiles.

Required Equipment and Materials:

- Compatible RT-PCR Devices*
Biorad CFX96 and Opus, Qiagene Rotorgene, Quantstudio Real-Time PCR Systems, Roche LightCycler 480 & 96, Mic qPCR Cycler, Corbett Realtime PCR, Slan Realtime PCR, Molarray MA6000, Himedia Insta Q96, Q48 and Q48M
- DNA Isolation (Manual or automated system)
- Vortex and Centrifuge
- Pipette Set (0.5-10µl and 10-100µl)
- Sterile Pipette Tips
- Cold Block
- PCR tubes or plates compatible with the respective Real-Time PCR Device
- +4°C Refrigerator
- Sterile Gloves

* Please contact for other compatible devices

Storage Conditions

- All components of the kit should be stored at -20°C until the expiration date.
- Kit components should undergo freeze-thaw cycles a maximum of 5 times. If more than 5 freeze-thaw cycles are anticipated, it is necessary to divide the kit components into smaller volumes for storage.

Experimental Preparation

Preparation of Materials:

- Materials stored at -20°C should be thawed on ice or cooling rack.
- Once thawed, liquid materials should be mixed well by pipetting.

Preparation of Reactions:

- Preparation of 6 tubes is required for each sample. Reaction volumes are specified in the table. The same tubes should be prepared for the NTC.

Content	Amount (µl)
PCR Master Mix	18
DNA Isolate / NTC	2
TOTAL	20

Real – Time PCR Protocol

Cycle	Temperature (°C)	Time (Seconds)	Fluorescent Dye
1	95 °C	300 s	
50	95 °C	10 s	
	60 °C	45 s	FAM, HEX, ROX, CY5
1	95 °C	180 s	
	55 °C	60 s	
Melt Analysis			
1	35 °C	1 s	
Initial Temp. (°C)	35 °C	Ramp Rate (°C /sec)	0.3
End Temperature	85 °C	Hold (sec)	1 s

Data Analysis – Interpretation of Results

TUBE 1				
Factor V LEIDEN (G1691A)	Normal (68°C)	Mutant (64°C)	Result	Emission Channel
	+	-	Normal	FAM
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (70°C)	Mutant (66°C)	Result	Emission Channel
Phrotrombin Factor II (G20210A)	+	-	Normal	HEX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (69°C)	Mutant (63°C)	Result	Emission Channel
MTHFR (C677T)	+	-	Normal	ROX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (63°C)	Mutant (70°C)	Result	Emission Channel
MTHFR (A1298C)	+	-	Normal	CY5
	-	+	Hom. Mutant	
	+	+	Heterozygous	
TUBE 2				
Factor XIII (V34L)	Normal (71°C)	Mutant (66°C)	Result	Emission Channel
	+	-	Normal	ROX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	4G (70°C)	5G (67°C)	Result	Emission Channel
PAI (4G/5G)	+	-	4G-4G	CY5
	-	+	5G-5G	
	+	+	4G—5G	

TUBE 3				
FGB (-455 G-A)	Normal (57°C)	Mutant (50°C)	Result	Emission Channel
	+	-	Normal	ROX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (67°C)	Mutant (63°C)	Result	Emission Channel
HPA1 (L33P)	+	-	Normal	CY5
	-	+	Hom. Mutant	
	+	+	Heterozygous	

TUBE 4				
H1299R (c.3980A>G)	Normal (69°C)	Mutant (63°C)	Result	Emission Channel
	+	-	Normal	ROX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (67°C)	Mutant (64°C)	Result	Emission Channel
FV CAMB (c.1001G>C)	+	-	Normal	CY5
	-	+	Hom. Mutant	
	+	+	Heterozygous	

TUBE 5				
APOB (R3500Q)	Normal (67°C)	Mutant (62°C)	Result	Emission Channel
	+	-	Normal	HEX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	Normal (70°C)	Mutant (63°C)	Result	Emission Channel
LTA	+	-	Normal	ROX
	-	+	Hom. Mutant	
	+	+	Heterozygous	
	DEL (67°C)	INS (62°C)	Result	Emission Channel
ACE (INS/DEL)	+	-	DEL-DEL	CY5
	-	+	INS-INS	
	+	+	INS-DEL	

TUBE 6				
APOE	RS429358 (ROX)		RS7412 (CY5)	
E2,E2	55°C		64°C	
E3,E3	55°C		71°C	
E4,E4	67°C		71°C	
E2,E3	55°C		71°C	64°C
E2,E4	55°C	67°C	71°C	64°C
E3,E4	55°C	67°C	71°C	

- After the Real-Time PCR stage, analysis begins by selecting the Melt Analysis option.
- Each tube must be analyzed separately.
- The analysis should be performed based on the presence or absence of peaks at the specified temperatures for each parameter.
- OctoClario™ Analysis Software can be used for automated analysis. Refer to the Software User Guide for program details.

Troubleshooting

1. If no melting peaks are observed in both controls and samples,
 - *Possible causes:* incomplete homogenization of the master mix; annealing/extension temperature set too high; insufficient or degraded template DNA.
 - *Solutions:* pipette and briefly centrifuge each reagent tube before setup; verify thermal-cycler temperatures strictly match IFU; repeat DNA extraction and check the quality of extracted DNA by fluorometry.
2. If control or sample melt-curve peaks are markedly lower in fluorescence than expected,
 - *Possible causes:* dye depletion due to ambient light exposure or prolonged room-temperature storage; expired or improperly stored reagents.
 - *Solutions:* protect master mix and reaction plates from light and keep on ice until use; check kit lot's expiration and storage logs; replace with a fresh kit if needed.
3. If melt peaks appear at temperatures outside the ± 1 °C window of the expected T_m ,
 - *Possible causes:* incorrect ramp rate or hold time during melt acquisition; nonspecific amplicons (e.g. primer-dimers).
 - *Solutions:* confirm melt-curve program parameters exactly match IFU.
4. If melt-curve derivative baselines are noisy or drift upward,
 - *Possible causes:* insufficient calibration of the instrument's optical modules; wells at plate edges subject to temperature lag.
 - *Solutions:* perform calibration with manufacturer's reference plate; avoid using outermost wells for critical controls or include a software-specified "edge-correction."
5. If sample genotyping calls fail QC despite clean control results,
 - *Possible causes:* sample inhibitors (e.g. heme, guanidine) carried over from extraction; pipetting errors.

- *Solutions:* repeat extraction using a spin-column kit with an inhibitor-removal step; employ multichannel pipette with filtered tips and include replicate wells.
6. If unexpected peaks appear in the no-template control (NTC),
 - *Possible causes:* airborne DNA contamination.
 - *Solutions:* enforce unidirectional workflow (pre-PCR vs. post-PCR areas).

Tip: Always consult your instrument's maintenance log and ensure that filter sets and heating blocks are within specification before attributing anomalies to the chemistry.

Warnings and Precautions

This test is for in vitro diagnostic purposes only and should be used strictly in accordance with the instructions provided by trained personnel. Clinical samples should be considered as materials potentially carrying an infection risk and should be prepared in a laminar flow cabinet.

- **Do not use expired kit.**
- **Avoid repeated freezing and thawing of reagents as it may reduce the sensitivity of the test.**
- **Ensure that reagents are thawed before use, then gently mix the tubes and centrifuge briefly.**
- **Prepare the reaction mixture quickly on ice or a cold block.**
- **Establish two separate work areas, one for isolation and the other for PCR.**
- **Avoid transferring pipettes, tubes, and other working materials between the work areas.**
- **Always use filtered sterile pipette tips.**
- **Use separate lab coats and gloves for each work area.**

Symbols Used

The following symbols used on labels and packaging of this product conform to the harmonized standard **TS EN ISO 15223-1**



: **Number of Reactions**



: **Expiration Date**



: **Lot Number**



: **Reference Number**



: **Temperature Limit**



: **Refer to the User Manual**



: **Manufacturer**



: **Keep Away From Direct Sunlight**

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